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ACR–SPR–SSR PRACTICE PARAMETER FOR THE PERFORMANCE AND INTERPRETATION OF MAGNETIC RESONANCE IMAGING (MRI) OF BONE AND SOFT TISSUE TUMORS

PREAMBLE

This document is an educational tool designed to assist practitioners in providing appropriate radiologic care for patients. Practice Parameters and Technical Standards are not inflexible rules or requirements of practice and are not intended, nor should they be used, to establish a legal standard of care¹. For these reasons and those set forth below, the American College of Radiology and our collaborating medical specialty societies caution against the use of these documents in litigation in which the clinical decisions of a practitioner are called into question.

The ultimate judgment regarding the propriety of any specific procedure or course of action must be made by the practitioner in light of all the circumstances presented. Thus, an approach that differs from the guidance in this document, standing alone, does not necessarily imply that the approach was below the standard of care. To the contrary, a conscientious practitioner may responsibly adopt a course of action different from that set forth in this document when, in the reasonable judgment of the practitioner, such course of action is indicated by the condition of the patient, limitations of available resources, or advances in knowledge or technology subsequent to publication of this document. However, a practitioner who employs an approach substantially different from the guidance in this document is advised to document in the patient record information sufficient to explain the approach taken.

The practice of medicine involves not only the science, but also the art of dealing with the prevention, diagnosis, alleviation, and treatment of disease. The variety and complexity of human conditions make it impossible to always reach the most appropriate diagnosis or to predict with certainty a particular response to treatment. Therefore, it should be recognized that adherence to the guidance in this document will not assure an accurate diagnosis or a successful outcome. All that should be expected is that the practitioner will follow a reasonable course of action based on current knowledge, available resources, and the needs of the patient to deliver effective and safe medical care. The sole purpose of this document is to assist practitioners in achieving this objective.

¹ *Iowa Medical Society and Iowa Society of Anesthesiologists v. Iowa Board of Nursing*, ___ N.W.2d ___ (Iowa 2013) Iowa Supreme Court refuses to find that the *ACR Technical Standard for Management of the Use of Radiation in Fluoroscopic Procedures* (Revised 2008) sets a national standard for who may perform fluoroscopic procedures in light of the standard's stated purpose that ACR standards are educational tools and not intended to establish a legal standard of care. See also, *Stanley v. McCarver*, 63 P.3d 1076 (Ariz. App. 2003) where in a concurring opinion the Court stated that "published standards or guidelines of specialty medical organizations are useful in determining the duty owed or the standard of care applicable in a given situation" even though ACR standards themselves do not establish the standard of care.

I. INTRODUCTION

This parameter was developed and written collaboratively by the American College of Radiology (ACR), the Society for Pediatric Radiology (SPR), and the Society of Skeletal Radiology (SSR).

Magnetic resonance imaging (MRI) is a proven and well-established imaging modality in the detection, evaluation, assessment, staging, and follow-up of tumors of the musculoskeletal system. Properly performed and interpreted, MRI not only contributes to initial diagnosis and identification of local recurrence but also serves as an important guide to biopsy and treatment planning as well as response to therapy. However, MRI of a tumor or suspected mass should be performed only for a valid medical reason and after careful consideration of alternative imaging modalities. An analysis of the strengths of MRI and other modalities should be weighed against their suitability for particular patients and particular clinical conditions. Radiographs should be used for the initial diagnosis of primary bone tumors. In addition, radiographs are usually the first imaging test performed for suspected soft-tissue masses and are particularly valuable for showing the presence and character of calcification, fat, or other radiopaque material. Radionuclide bone scanning and single-photon emission computed tomography (SPECT), with or without CT coregistration, is often used when occult bone disease is suspected, or to screen the entire skeleton for conditions such as metastases. Other nuclear medicine examinations have a role for specific clinical scenarios (eg, a labeled white blood cell study for suspected osteomyelitis). Computed tomography (CT) shows detailed bone anatomy and aids in identifying osteoid and chondroid matrix. Sonography may aid in examination of soft-tissue masses (eg, cystic versus solid, assessment of vascularity) [1,2]. Conventional, MR, or CT angiography remains useful for evaluating tumor vascularity, identifying the relationship of the lesion to adjacent major blood vessels, planning resection and reconstruction, and providing access for presurgical embolization [3]. Positron emission tomography (with or without CT coregistration) can help stage and grade tumors [7-12], assess response to therapy [13-16], and detect tumor recurrence [10,17], but it may not reliably discriminate between benign and malignant tumors [8,18].

Although MRI is one of the most sensitive, noninvasive diagnostic tests for detecting anatomic abnormalities of the musculoskeletal system, findings may be misleading if not closely correlated with radiographs, clinical history, clinical examination, and physiologic tests [19,20]. Adherence to the following guidelines will enhance the probability of detecting such abnormalities.

II. INDICATIONS

Indications for MRI of soft-tissue and bone tumors include, but are not limited to, the following:

1. Initial characterization, detection, or exclusion of tumors [21-36]
2. Local staging of tumors [37-41]
3. Evaluation of tumors prior to biopsy, surgery, chemotherapy, and/or radiotherapy [4,6,29,37,42-44]
4. Evaluation of the response of tumors to treatment, including neoadjuvant chemotherapy, postresection chemotherapy, and radiation therapy [45-56]
5. Detection and evaluation of complications related to tumors or their treatment, including hemorrhage, infection, and neurologic and vascular conditions [29,52,55-65]
6. Post-treatment and long-term surveillance and characterization of local, regional, and distant tumor recurrences [53,54]

III. QUALIFICATIONS AND RESPONSIBILITIES OF PERSONNEL

See the [ACR Practice Parameter for Performing and Interpreting Magnetic Resonance Imaging \(MRI\)](#) [66].

IV. SAFETY GUIDELINES AND POSSIBLE CONTRAINDICATIONS

See the [ACR Practice Parameter for Performing and Interpreting Magnetic Resonance Imaging \(MRI\)](#) [66], the [ACR Guidance Document on MR Safe Practices](#) [67], and the [ACR Manual on Contrast Media](#) [68].

Peer-reviewed literature pertaining to MR safety should be reviewed on a regular basis [69,70].

V. SPECIFICATIONS OF THE EXAMINATION

The supervising physician must have complete understanding of the indications, risks, and benefits of the examination, as well as alternative imaging procedures. The physician must be familiar with potential hazards associated with MRI, including potential adverse reactions to contrast media. The physician should be familiar with relevant ancillary studies that the patient may have undergone. The physician performing MRI interpretation must have a clear understanding and knowledge of the anatomy and pathophysiology relevant to the MRI examination.

The written or electronic request for MRI of bone and soft tissue tumors should provide sufficient information to demonstrate the medical necessity of the examination and allow for its proper performance and interpretation.

Documentation that satisfies medical necessity includes 1) signs and symptoms and/or 2) relevant history (including known diagnoses). Additional information regarding the specific reason for the examination or a provisional diagnosis would be helpful and may at times be needed to allow for the proper performance and interpretation of the examination.

The request for the examination must be originated by a physician or other appropriately licensed health care provider. The accompanying clinical information should be provided by a physician or other appropriately licensed health care provider familiar with the patient's clinical problem or question and consistent with the state's scope of practice requirements. (ACR Resolution 35, adopted in 2006)

The supervising physician must also understand the pulse sequences to be used and their effect on the appearance of the images, including the potential generation of image artifacts. Standard imaging protocols may be established and varied on a case-by-case basis when necessary. These protocols should be reviewed and updated periodically.

A. Patient Selection

The physician responsible for the examination should supervise patient selection and preparation, and be available in person or by phone for consultation. Patients must be screened and interviewed prior to the examination to exclude individuals who may be at risk by exposure to the MR environment.

Certain indications require administration of intravenous (IV) contrast media. IV contrast enhancement should be performed using appropriate injection protocols and in accordance with the institution's policy on IV contrast utilization (See the [ACR-SPR Practice Parameter for the Use of Intravascular Contrast Media](#) [71]).

Pediatric patients or patients suffering from anxiety or claustrophobia may require sedation or additional assistance. Administration of moderate sedation or general anesthesia may be needed to achieve a successful examination, particularly in young children. If moderate sedation is necessary, refer to the [ACR-SIR Practice Parameter for Sedation/Analgesia](#) [72]. Young children may require sedation or general anesthesia in order to prevent patient motion during the MR examination.

B. Facility Requirements

Appropriate emergency equipment and medications must be immediately available to treat adverse reactions associated with administered medications. The equipment and medications should be monitored for inventory and drug expiration dates on a regular basis. The equipment, medications, and other emergency support must also be appropriate for the range of ages and sizes in the patient population.

C. Examination Technique

Diagnostic quality MRI of suspected bone and soft-tissue masses can be performed using a variety of magnetic designs (closed-bore whole body, open whole body) and a variety of field strengths [23,25,28,31]. Regardless of system design, efforts should be made to maximize signal-to-noise ratios. Field of view (FOV) should be tailored to the size of the patient and the size of the suspected mass [25,63,73,74]. For example, a 48-cm FOV would be appropriate for an extremely large tumor of the pelvis or thigh, whereas a 12-cm FOV may be appropriate for a small mass in the foot. At times, additional sequences with a larger FOV will be necessary to evaluate proximal or distal spread of disease. It is important to obtain as many transverse, sagittal, or coronal images through the lesion as is reasonable. Slice thicknesses will also vary depending on the size of the lesion [25]. For example, a 1-cm mass might require 3-mm thick slices, whereas a tumor greater than 30 cm in size may be appropriately imaged with 1-cm slice thickness [25]. An interslice gap may be chosen to decrease signal loss due to cross-talk [73] but in general should be no more than one-half of the slice width and should not impair complete visualization of the mass. The imaging matrix should balance the intravoxel signal-to-noise ratio with desired in-plane spatial resolution.

The size and location of the lesion will often dictate the most appropriate coil to use for imaging. Small lesions or lesions located in the extremities will often be best imaged using a local surface coil, a cylindrical coil, or a dedicated joint coil. For extremely large lesions or lesions involving the torso, the body coil may be a more appropriate choice [25,41,44]. Every attempt should be made to include the entire soft-tissue or bone tumor and associated edema in the imaged volume. Additionally, for high-grade sarcomas of bone, the entire bone should be imaged to evaluate for skip lesions and regional metastases. The use of a multiple-channel receiver coil unit may allow the use of parallel imaging techniques to reduce overall scan time or improve signal-to-noise ratio and may be useful in reducing motion-related artifacts [75,76].

For patients with more than one suspected bone or soft-tissue mass it may be necessary to perform separate MR examinations. For example, a patient with a mass involving both the pelvis and leg may require 2 separate studies.

When an MR imager of field strength less than 1.5T is used to image bone and soft-tissue tumors, then other imaging parameters—such as the receiver bandwidth and number of acquisitions—will require modification to ensure adequate spatial and contrast resolution for confident diagnosis, often at the expense of longer examination times [63,77]. It may also be more difficult to achieve uniform fat suppression on low-field systems using spectrally selective RF presaturation pulses, potentially necessitating the use of Dixon or short TI inversion recovery (STIR) techniques [78-81]. Other systems may be more prone to imaging artifacts (eg, chemical shift artifact on high-field magnets), again necessitating modification of imaging parameters such as receiver bandwidth to ensure that these artifacts do not detract from the diagnostic quality of the resultant images. Some MRI systems may not be appropriate for specific indications. For example, high-resolution evaluation of a small mass may not be feasible with a low-field, open magnet, regardless of the chosen imaging parameters [82].

MR imaging of bone and soft-tissue tumors usually includes images in at least 2 orthogonal planes (transverse, sagittal, and coronal) [23,25,26,32,63]. The long axis images may be oriented orthogonal to the magnetic bore or may be angled to better identify specific anatomic structures. The coverage of the tumor ideally should include all of the anterior, posterior, medial, lateral, superior, and inferior margins of the mass [6,23,25].

MRI of suspected bone and soft-tissue tumors can be performed with a variety of pulse sequences. The choice of sequences can be tailored to optimize the examination for specific clinical questions and according to local preferences. An imaging protocol would usually be composed of at least one T1-weighted image and one fluid-sensitive T2-weighted sequence with or without fat suppression.

Short-TE images with a relatively short TR (T1-weighted) are commonly used to evaluate tumors [23,25,73,77]. Because of the image blurring inherent in a fast spin-echo image made with a short effective TE, conventional spin-echo imaging may be preferred [23,25,73,77]. Properly optimized, however, some investigators have used fast spin-echo imaging for T1-weighted images. To demonstrate pathologic tissues, T2-weighted (fluid-sensitive) imaging using conventional spin-echo or fast spin-echo sequences are most commonly used [78-81,83]. T1-weighted spoiled gradient-echo chemical shift imaging (ie, water-fat in-phase/opposed-phase imaging) can be used to demonstrate the presence of lipid components in tissues and may help discriminate benign from malignant disease processes such as in evaluation of fractures and bone marrow infiltration [84,85]. Gradient-recalled

sequences may also be valuable, in particular in evaluating for internal areas of hemorrhage, gas, ossification, or calcification.

T1-weighted sequences are routinely done without fat suppression to depict anatomic relationships; however, the addition of fat suppression may be helpful to detect hemorrhage or fat within a mass and when intravenous contrast is given [86]. Fluid-sensitive images, obtained with long TR using conventional or fast spin-echo sequences, can be used to characterize bone and soft-tissue tumors, providing complementary information to the T1-weighted images. Therefore, a combination of both T1-weighted and T2-weighted images is typically performed in each imaging plane [23,79-81,83]. Lesion conspicuity may be increased with the addition of fat suppression to fluid-sensitive images; however, fat-suppressed imaging decreases the variation in tumor signal intensities that may be useful in tissue characterization. T2-weighted sequences can be performed with or without fat suppression, or STIR (Short Tau Inversion Recovery) sequences can be used [79,80,83]. A combination of techniques may prove advantageous. For example, the transverse images may be obtained without fat suppression and the long axis planes (sagittal and/or coronal images) performed with fat suppression or STIR sequences. The exact TR, TE, and flip angle chosen will depend on the field strength of the magnet and the relative contrast weighting desired [87-89].

Various techniques may be used to minimize the MR artifacts that can reduce imaging quality. Wraparound artifact, including that originating from signal received from other parts of the body, can be reduced by phase using oversampling, by switching the phase and frequency readout directions, by presaturation pulses, or by using radiofrequency shielding. Truncation (Gibbs) artifacts may obscure or mimic intralesional detail and can be reduced by changing the phase-encoding direction. Involuntary patient motion is best controlled by ensuring patient comfort combined with gentle immobilization or sedation when necessary and often requires sedation or general anesthesia for young children [63,90]. Use of MR systems and coils that provide a high signal-to-noise ratio, such as high-field (3T) MR systems and multichannel coils, with or without parallel imaging, can reduce overall scan duration and individual sequence scan times and may help reduce bulk motion artifacts and patient discomfort [75,76]. Flowing blood can produce ghosting artifacts, which can be reduced with presaturation pulses or the use of gradient moment nulling [63,90].

In many cases it may be advantageous to administer a gadolinium-based intravenous contrast agent [91-97]. Intravenous contrast may be helpful to differentiate cysts from solid masses and may provide additional details of the imaging features of bone and soft-tissue masses [83,92,93]. Subtracting the precontrast images from the postcontrast images may be beneficial to show subtle areas of enhancement and to distinguish enhancement from adjacent fat or hemorrhage [98]. Fast, multiphase dynamic contrast-enhanced imaging can provide analysis of tumor perfusion kinetics, including parametric perfusion data, that may help to distinguish malignant from benign tumors [99-101], to stage tumors and response to therapy [49,102-104], to determine an optimal site for biopsy [104], or to improve tumor detection or evaluate potential extension of tumor cells along related fascial planes [105]. The decision to use intravenous contrast should be based on medical appropriateness.

Follow-up MR imaging of musculoskeletal tumors is generally performed using sequences similar to those used for initial diagnosis, including T1-weighted and T2-weighted images [53,54]. Follow-up MR examinations of patients with previously treated soft-tissue tumors often benefit by the addition of intravenous gadolinium chelates [52,53]. Protocols for follow-up and interpretation of MRI findings vary depending on the type of tumor, the therapeutic methods used, and the aggressiveness of the tumor (See the [ACR Appropriateness Criteria® Follow-up of Malignant or Aggressive Musculoskeletal Tumors](#) [106]).

MR spectroscopy may be useful in gauging therapy response and tumor staging [113-118]. It may also be used to detect certain metabolites in tumors to help in lesion characterization [115,119-124], but caution should be used in interpretation as some metabolites that were thought to be specific may not be (eg, choline for malignant tumors [125]). New imaging sequences employing isotropic or near-isotropic 3-D sequences (eg, IDEAL, Space, Cubem, etc) can produce images with shorter scan duration but have not been evaluated for imaging of musculoskeletal tumors at this time. Whole-body MR screening examinations can be useful both for staging of disseminated or

hematologic tumors such as multiple myeloma and to limit radiation dose to pediatric and pregnant patients [126-130].

For interpretation, the images can be printed on film or viewed on a workstation. If hardcopy viewing is used, some practices may film the images with magnified or narrowed window settings, but this can be left to local preferences. MR examinations in patients with suspected tumors should be read cautiously and preferably in conjunction with available radiographs. Since local recurrence may often appear similar to the original tumor, MR imaging following treatment or surgery should ideally be interpreted with comparison to prior MRI exams, including the preoperative or pretreatment MRI, if available. There are many pitfalls and artifacts that can suggest that a non-neoplastic mass is an aggressive tumor or that a malignant tumor appears to be a benign lesion based on the MR appearance alone [83,131,132]. Furthermore, imaging artifacts can also contribute to incorrect staging of tumors [83,131,132].

VI. DOCUMENTATION

Reporting should be in accordance with the [ACR Practice Parameter for Communication of Diagnostic Imaging Findings](#) [133].

The report should address the presence or absence of a mass, the size of the lesion and its composition (hemorrhage, necrosis, etc), signal intensity, and enhancement characteristics. When imaging is sufficiently characteristic, a diagnosis or differential diagnosis should be provided. A description of the anatomic location of a tumor, including its intracompartmental and extracompartmental extent, as well as its relationships to adjacent major muscles, vessels, and nerves, will contribute to the tumor's grading and staging. The presence or absence of fascial extension of tumor should be described, which will contribute to the surgical resection planning. The presence or absence of any regional lymphadenopathy or skip lesions should be noted.

VII. EQUIPMENT SPECIFICATIONS

The MRI equipment specifications and performance must meet all state and federal requirements. The requirements include, but are not limited to, specifications of maximum static magnetic strength, maximum rate of change of the magnetic field strength (dB/dt), maximum radiofrequency power deposition (specific absorption rate), and maximum acoustic noise levels.

VIII. QUALITY CONTROL AND IMPROVEMENT, SAFETY, INFECTION CONTROL, AND PATIENT EDUCATION

Policies and procedures related to quality, patient education, infection control, and safety should be developed and implemented in accordance with the ACR Policy on Quality Control and Improvement, Safety, Infection Control, and Patient Education appearing under the heading *Position Statement on QC & Improvement, Safety, Infection Control, and Patient Education* on the ACR website (<http://www.acr.org/guidelines>).

Specific policies and procedures related to MRI safety should be in place with documentation that is updated annually and compiled under the supervision and direction of the supervising MRI physician. Guidelines should be provided that deal with potential hazards associated with MRI examination to the patient as well as to others in the immediate area [69,70,134]. Screening forms must also be provided to detect those patients who may be at risk for adverse events associated with the MRI examination [69,70,134,135].

Equipment monitoring should be in accordance with the [ACR-AAPM Technical Standard for Diagnostic Medical Physics Performance Monitoring of Magnetic Resonance Imaging \(MRI\) Equipment](#) [136].

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REFERENCES

1. Pathria MN, Zlatkin M, Sartoris DJ, Scheible W, Resnick D. Ultrasonography of the popliteal fossa and lower extremities. *Radiol Clin North Am.* 1988;26(1):77-85.
2. Taylor GA, Perlman EJ, Scherer LR, Gearhart JP, Leventhal BG, Wiley J. Vascularity of tumors in children: evaluation with color Doppler imaging. *AJR Am J Roentgenol.* 1991;157(6):1267-1271.
3. Lois JF, Fischer HJ, Deutsch LS, Stambuk EC, Gomes AS. Angiography in soft tissue sarcomas. *Cardiovasc Intervent Radiol.* 1984;7(6):309-316.
4. Feydy A, Anract P, Tomeno B, Chevrot A, Drape JL. Assessment of vascular invasion by musculoskeletal tumors of the limbs: use of contrast-enhanced MR angiography. *Radiology.* 2006;238(2):611-621.
5. Lang P, Grampp S, Vahlensieck M, et al. Primary bone tumors: value of MR angiography for preoperative planning and monitoring response to chemotherapy. *AJR Am J Roentgenol.* 1995;165(1):135-142.
6. Swan JS, Grist TM, Sproat IA, Heiner JP, Wiersma SR, Heisey DM. Musculoskeletal neoplasms: preoperative evaluation with MR angiography. *Radiology.* 1995;194(2):519-524.
7. Eary JF, O'Sullivan F, Powitan Y, et al. Sarcoma tumor FDG uptake measured by PET and patient outcome: a retrospective analysis. *Eur J Nucl Med Mol Imaging.* 2002;29(9):1149-1154.
8. Ioannidis JP, Lau J. 18F-FDG PET for the diagnosis and grading of soft-tissue sarcoma: a meta-analysis. *J Nucl Med.* 2003;44(5):717-724.
9. Lisle JW, Eary JF, O'Sullivan J, Conrad EU. Risk assessment based on FDG-PET imaging in patients with synovial sarcoma. *Clin Orthop Relat Res.* 2009;467(6):1605-1611.
10. Lucas JD, O'Doherty MJ, Wong JC, et al. Evaluation of fluorodeoxyglucose positron emission tomography in the management of soft-tissue sarcomas. *J Bone Joint Surg Br.* 1998;80(3):441-447.
11. Tateishi U, Yamaguchi U, Seki K, Terauchi T, Arai Y, Kim EE. Bone and soft-tissue sarcoma: preoperative staging with fluorine 18 fluorodeoxyglucose PET/CT and conventional imaging. *Radiology.* 2007;245(3):839-847.
12. Volker T, Denecke T, Steffen I, et al. Positron emission tomography for staging of pediatric sarcoma patients: results of a prospective multicenter trial. *J Clin Oncol.* 2007;25(34):5435-5441.
13. Kasper B, Dietrich S, Dimitrakopoulou-Strauss A, et al. Early prediction of therapy outcome in patients with high-risk soft tissue sarcoma using positron emission tomography. *Onkologie.* 2008;31(3):107-112.
14. Schuetze SM, Rubin BP, Vernon C, et al. Use of positron emission tomography in localized extremity soft tissue sarcoma treated with neoadjuvant chemotherapy. *Cancer.* 2005;103(2):339-348.
15. Vernon CB, Eary JF, Rubin BP, Conrad EU, 3rd, Schuetze S. FDG PET imaging guided re-evaluation of histopathologic response in a patient with high-grade sarcoma. *Skeletal Radiol.* 2003;32(3):139-142.
16. Ye Z, Zhu J, Tian M, et al. Response of osteogenic sarcoma to neoadjuvant therapy: evaluated by 18F-FDG- PET. *Ann Nucl Med.* 2008;22(6):475-480.
17. Johnson GR, Zhuang H, Khan J, Chiang SB, Alavi A. Roles of positron emission tomography with fluorine-18-deoxyglucose in the detection of local recurrent and distant metastatic sarcoma. *Clin Nucl Med.* 2003;28(10):815-820.

18. Strobel K, Bode B, Lardinois D, Exner U. PET-positive fibrous dysplasia--a potentially misleading incidental finding in a patient with intimal sarcoma of the pulmonary artery. *Skeletal Radiol.* 2007;36 Suppl 1:S24-28.
19. Oliveira AM, Nascimento AG. Grading in soft tissue tumors: principles and problems. *Skeletal Radiol.*2001;30(10):543-559.
20. Stacy GS, Mahal RS, Peabody TD. Staging of bone tumors: a review with illustrative examples. *AJR Am J Roentgenol.* 2006;186(4):967-976.
21. Aboulafia AJ, Kennon RE, Jelinek JS. Benign bone tumors of childhood. *J Am Acad Orthop Surg.* 1999;7(6):377-388.
22. Dalinka MK, Zlatkin MB, Chao P, Kricun ME, Kressel HY. The use of magnetic resonance imaging in the evaluation of bone and soft-tissue tumors. *Radiol Clin North Am.* 1990;28(2):461-470.
23. Hagggar AM, Froelich JW. MR imaging strategies in primary and metastatic malignancy. *Radiol Clin North Am.* 1988;26(3):689-696.
24. Hoffer FA. Primary skeletal neoplasms: osteosarcoma and ewing sarcoma. *Top Magn Reson Imaging.* 2002;13(4):231-239.
25. Kransdorf MJ, Bancroft LW, Peterson JJ, Murphey MD, Foster WC, Temple HT. Imaging of fatty tumors: distinction of lipoma and well-differentiated liposarcoma. *Radiology.* 2002;224(1):99-104.
26. Kransdorf MJ, Jelinek JS, Moser RP, Jr., et al. Soft-tissue masses: diagnosis using MR imaging. *AJR Am J Roentgenol.* 1989;153(3):541-547.
27. Ma LD. Magnetic resonance imaging of musculoskeletal tumors: skeletal and soft tissue masses. *Curr Probl Diagn Radiol.* 1999;28(2):29-62.
28. Murphey MD, Gross TM, Rosenthal HG, Neff JR. Magnetic resonance imaging of soft tissue and cystic masses about the knee. *Top Magn Reson Imaging.* 1993;5(4):263-282.
29. Nomikos GC, Murphey MD, Kransdorf MJ, Bancroft LW, Peterson JJ. Primary bone tumors of the lower extremities. *Radiol Clin North Am.* 2002;40(5):971-990.
30. Pettersson H, Gillespy T, 3rd, Hamlin DJ, et al. Primary musculoskeletal tumors: examination with MR imaging compared with conventional modalities. *Radiology.* 1987;164(1):237-241.
31. Ritchie DA, Davies AM. MR imaging of tumors and tumor-like lesions of the shoulder girdle. *Magn Reson Imaging Clin N Am.* 2004;12(1):125-141, vii.
32. Sundaram M. Magnetic resonance imaging for solitary lesions of bone: when, why, how useful? *J Orthop Sci.* 1999;4(5):384-396.
33. Sundaram M, McGuire MH. Computed tomography or magnetic resonance for evaluating the solitary tumor or tumor-like lesion of bone? *Skeletal Radiol.* 1988;17(6):393-401.
34. Sundaram M, McGuire MH, Herbold DR. Magnetic resonance imaging of soft tissue masses: an evaluation of fifty-three histologically proven tumors. *Magn Reson Imaging.* 1988;6(3):237-248.
35. Sundaram M, McLeod RA. MR imaging of tumor and tumorlike lesions of bone and soft tissue. *AJR Am J Roentgenol.* 1990;155(4):817-824.
36. Tehranzadeh J, Mnaymneh W, Ghavam C, Morillo G, Murphy BJ. Comparison of CT and MR imaging in musculoskeletal neoplasms. *J Comput Assist Tomogr.* 1989;13(3):466-472.
37. Bloem JL, Taminiau AH, Eulerink F, Hermans J, Pauwels EK. Radiologic staging of primary bone sarcoma: MR imaging, scintigraphy, angiography, and CT correlated with pathologic examination. *Radiology.* 1988;169(3):805-810.
38. Demas BE, Heelan RT, Lane J, Marcove R, Hajdu S, Brennan MF. Soft-tissue sarcomas of the extremities: comparison of MR and CT in determining the extent of disease. *AJR Am J Roentgenol.* 1988;150(3):615-620.
39. Frank JA, Ling A, Patronas NJ, et al. Detection of malignant bone tumors: MR imaging vs scintigraphy. *AJR Am J Roentgenol.* 1990;155(5):1043-1048.
40. Panicek DM, Gatsonis C, Rosenthal DI, et al. CT and MR imaging in the local staging of primary malignant musculoskeletal neoplasms: Report of the Radiology Diagnostic Oncology Group. *Radiology.*1997;202(1):237-246.
41. Saifuddin A. The accuracy of imaging in the local staging of appendicular osteosarcoma. *Skeletal Radiol.* 2002;31(4):191-201.
42. Elias DA, White LM, Simpson DJ, et al. Osseous invasion by soft-tissue sarcoma: assessment with MR imaging. *Radiology.* 2003;229(1):145-152.

43. Enneking WF, Spanier SS, Goodman MA. A system for the surgical staging of musculoskeletal sarcoma. *Clin Orthop Relat Res.* 1980(153):106-120.
44. Mouloupoulos LA, Dimopoulos MA, Vourtsi A, Gouliamos A, Vlahos L. Bone lesions with soft-tissue mass: magnetic resonance imaging diagnosis of lymphomatous involvement of the bone marrow versus multiple myeloma and bone metastases. *Leuk Lymphoma.* 1999;34(1-2):179-184.
45. Baur A, Stabler A, Wendtner CM, et al. MR-imaging changes of musculoskeletal soft-tissue sarcomas associated with neoadjuvant chemotherapy and hyperthermia. *Int J Hyperthermia.* 2003;19(4):391-401.
46. Bearcroft PW, Davies AM. Follow-up of musculoskeletal tumours. 2. Metastatic disease. *Eur Radiol.* 1999;9(2):192-200.
47. Biondetti PR, Ehman RL. Soft-tissue sarcomas: use of textural patterns in skeletal muscle as a diagnostic feature in postoperative MR imaging. *Radiology.* 1992;183(3):845-848.
48. Choi H, Varma DG, Fornage BD, Kim EE, Johnston DA. Soft-tissue sarcoma: MR imaging vs sonography for detection of local recurrence after surgery. *AJR Am J Roentgenol.* 1991;157(2):353-358.
49. Dyke JP, Panicek DM, Healey JH, et al. Osteogenic and Ewing sarcomas: estimation of necrotic fraction during induction chemotherapy with dynamic contrast-enhanced MR imaging. *Radiology.* 2003;228(1):271-278.
50. Fletcher BD. Response of osteosarcoma and Ewing sarcoma to chemotherapy: imaging evaluation. *AJR Am J Roentgenol.* 1991;157(4):825-833.
51. Reuther G, Mutschler W. Detection of local recurrent disease in musculoskeletal tumors: magnetic resonance imaging versus computed tomography. *Skeletal Radiol.* 1990;19(2):85-90.
52. van der Woude HJ, Bloem JL, Hogendoorn PC. Preoperative evaluation and monitoring chemotherapy in patients with high-grade osteogenic and Ewing's sarcoma: review of current imaging modalities. *Skeletal Radiol.* 1998;27(2):57-71.
53. Vanel D, Lacombe MJ, Couanet D, Kalifa C, Spielmann M, Genin J. Musculoskeletal tumors: follow-up with MR imaging after treatment with surgery and radiation therapy. *Radiology.* 1987;164(1):243-245.
54. Vanel D, Shapeero LG, De Baere T, et al. MR imaging in the follow-up of malignant and aggressive soft-tissue tumors: results of 511 examinations. *Radiology.* 1994;190(1):263-268.
55. Varma DG, Jackson EF, Pollock RE, Benjamin RS. Soft-tissue sarcoma of the extremities. MR appearance of post-treatment changes and local recurrences. *Magn Reson Imaging Clin N Am.* 1995;3(4):695-712.
56. Verstraete KL, Lang P. Post-therapeutic magnetic resonance imaging of bone tumors. *Top Magn Reson Imaging.* 1999;10(4):237-246.
57. Bush CH. The magnetic resonance imaging of musculoskeletal hemorrhage. *Skeletal Radiol.* 2000;29(1):1-9.
58. Fritz RC, Helms CA, Steinbach LS, Genant HK. Suprascapular nerve entrapment: evaluation with MR imaging. *Radiology.* 1992;182(2):437-444.
59. Kothari NA, Pelchovitz DJ, Meyer JS. Imaging of musculoskeletal infections. *Radiol Clin North Am.* 2001; 39(4):653-671.
60. Lenchik L, Dovgan DJ, Kier R. CT of the iliopsoas compartment: value in differentiating tumor, abscess, and hematoma. *AJR Am J Roentgenol.* 1994;162(1):83-86.
61. Panicek DM, Casper ES, Brennan MF, Hajdu SI, Heelan RT. Hemorrhage simulating tumor growth in malignant fibrous histiocytoma at MR imaging. *Radiology.* 1991;181(2):398-400.
62. Roebuck DJ. Skeletal complications in pediatric oncology patients. *Radiographics.* 1999;19(4):873-885.
63. Rubin DA, Kneeland JB. MR imaging of the musculoskeletal system: technical considerations for enhancing image quality and diagnostic yield. *AJR Am J Roentgenol.* 1994;163(5):1155-1163.
64. Struk DW, Munk PL, Lee MJ, Ho SG, Worsley DF. Imaging of soft tissue infections. *Radiol Clin North Am.* 2001;39(2):277-303.
65. Unger EC, Glazer HS, Lee JK, Ling D. MRI of extracranial hematomas: preliminary observations. *AJR Am J Roentgenol.* 1986;146(2):403-407.
66. American College of Radiology. ACR Practice Parameter for Performing and Interpreting Magnetic Resonance Imaging (MRI). 2014; <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/MR-Perf-Interpret.pdf>. Accessed October 8, 2014.

67. Kanal E, Barkovich AJ, Bell C, et al. ACR guidance document for safe MR practices: 2007. *AJR Am J Roentgenol.* 2007;188(6):1447-1474.
68. American College of Radiology. Manual on Contrast Media. 2009; http://www.acr.org/SecondaryMainMenuCategories/quality_safety/contrast_manual.aspx. Accessed September 11, 2009.
69. Shellock FG. *Reference Manual for Magnetic Resonance Safety, Implants and Devices.* 2009 ed. Los Angeles, Calif.: Biomedical Research Publishing Company; 2009.
70. Shellock FG, Crues JV. MR procedures: biologic effects, safety, and patient care. *Radiology.* 2004;232(3):635-652.
71. American College of Radiology. ACR–SPR Practice Parameter for the Use of Intravascular Contrast Media. 2014; <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/IVCM.pdf>. Accessed October 8, 2014.
72. American College of Radiology. ACR–SIR Practice Parameter for Sedation/Analgesia. 2014; <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/Sed-Analgesia.pdf>. Accessed October 8, 2014.
73. Kneeland JB, Shimakawa A, Wehrli FW. Effect of intersection spacing on MR image contrast and study time. *Radiology.* 1986;158(3):819-822.
74. Pettersson H, Slone RM, Spanier S, Gillespy T, 3rd, Fitzsimmons JR, Scott KN. Musculoskeletal tumors: T1 and T2 relaxation times. *Radiology.* 1988;167(3):783-785.
75. Jaramillo D, Laor T. Pediatric musculoskeletal MRI: basic principles to optimize success. *Pediatr Radiol.* 2008;38(4):379-391.
76. Romaneehsen B, Oberholzer K, Muller LP, Kreitner KF. Rapid musculoskeletal magnetic resonance imaging using integrated parallel acquisition techniques (IPAT)--initial experiences. *Rofo.* 2003;175(9):1193-1197.
77. Erickson SJ. High-resolution imaging of the musculoskeletal system. *Radiology.* 1997;205(3):593-618.
78. Delfaut EM, Beltran J, Johnson G, Rousseau J, Marchandise X, Cotten A. Fat suppression in MR imaging: techniques and pitfalls. *Radiographics.* 1999;19(2):373-382.
79. Mirowitz SA. Fast scanning and fat-suppression MR imaging of musculoskeletal disorders. *AJR Am J Roentgenol.* 1993;161(6):1147-1157.
80. Rybicki FJ, Chung T, Reid J, Jaramillo D, Mulkern RV, Ma J. Fast three-point dixon MR imaging using low-resolution images for phase correction: a comparison with chemical shift selective fat suppression for pediatric musculoskeletal imaging. *AJR Am J Roentgenol.* 2001;177(5):1019-1023.
81. Shuman WP, Baron RL, Peters MJ, Tazioli PK. Comparison of STIR and spin-echo MR imaging at 1.5 T in 90 lesions of the chest, liver, and pelvis. *AJR Am J Roentgenol.* 1989;152(4):853-859.
82. Ziedses des Plantes BG, Koster K. Comparison of low-field versus high-field MR imaging. *Eur J Radiol.* 1995;20(2):156-158.
83. Kransdorf MJ, Murphey, MD. *Imaging of Soft Tissue Tumors.* 2nd ed. Philadelphia, Pa.: Lippincott Williams & Wilkins; 2006.
84. Rosen BR, Fleming DM, Kushner DC, et al. Hematologic bone marrow disorders: quantitative chemical shift MR imaging. *Radiology.* 1988;169(3):799-804.
85. Zajick DC, Jr., Morrison WB, Schweitzer ME, Parellada JA, Carrino JA. Benign and malignant processes: normal values and differentiation with chemical shift MR imaging in vertebral marrow. *Radiology.* 2005;237(2):590-596.
86. Gielen JL, De Schepper AM, Parizel PM, Wang XL, Vanhoenacker F. Additional value of magnetic resonance with spin echo T1-weighted imaging with fat suppression in characterization of soft tissue tumors. *J Comput Assist Tomogr.* 2003;27(3):434-441.
87. Erickson SJ. High-resolution imaging of the musculoskeletal system. *Radiology.* 1997;205(9393511):593-618.
88. Hagggar AM, Froelich JW. MR imaging strategies in primary and metastatic malignancy. *Radiol Clin North Am.* 1988;26(3287441):689-696.
89. Rubin DA, Kneeland JB. MR imaging of the musculoskeletal system: technical considerations for enhancing image quality and diagnostic yield. *AJR Am J Roentgenol.* 1994;163(7976893):1155-1163.
90. Haacke EM, Lenz GW. Improving MR image quality in the presence of motion by using rephasing gradients. *AJR Am J Roentgenol.* 1987;148(6):1251-1258.
91. Benedikt RA, Jelinek JS, Kransdorf MJ, Moser RP, Berrey BH. MR imaging of soft-tissue masses: role of gadopentetate dimeglumine. *J Magn Reson Imaging.* 1994;4(3):485-490.

92. Erlemann R, Reiser MF, Peters PE, et al. Musculoskeletal neoplasms: static and dynamic Gd-DTPA--enhanced MR imaging. *Radiology*. 1989;171(3):767-773.
93. Kransdorf MJ, Murphey MD. The use of gadolinium in the MR evaluation of soft tissue tumors. *Semin Ultrasound CT MR*. 1997;18(4):251-268.
94. Mirowitz SA, Totty WG, Lee JK. Characterization of musculoskeletal masses using dynamic Gd-DTPA enhanced spin-echo MRI. *J Comput Assist Tomogr*. 1992;16(1):120-125.
95. Pettersson H, Eliasson J, Egund N, et al. Gadolinium-DTPA enhancement of soft tissue tumors in magnetic resonance imaging--preliminary clinical experience in five patients. *Skeletal Radiol*. 1988;17(5):319-323.
96. Shapeero LG, Vanel D, Verstraete KL, Bloem JL. Dynamic Contrast-Enhanced MR Imaging for Soft Tissue Sarcomas. *Semin Musculoskelet Radiol*. 1999;3(2):101-114.
97. Verstraete KL, Lang P. Bone and soft tissue tumors: the role of contrast agents for MR imaging. *Eur J Radiol*. 2000;34(3):229-246.
98. Hanna SL, Langston JW, Gronemeyer SA, Fletcher BD. Subtraction technique for contrast-enhanced MR images of musculoskeletal tumors. *Magn Reson Imaging*. 1990;8(3):213-215.
99. Tuncbilek N, Karakas HM, Okten OO. Dynamic contrast enhanced MRI in the differential diagnosis of soft tissue tumors. *Eur J Radiol*. 2005;53(3):500-505.
100. van der Woude HJ, Verstraete KL, Hogendoorn PC, Taminiau AH, Hermans J, Bloem JL. Musculoskeletal tumors: does fast dynamic contrast-enhanced subtraction MR imaging contribute to the characterization? *Radiology*. 1998;208(3):821-828.
101. van Rijswijk CS, Geirnaerd MJ, Hogendoorn PC, et al. Soft-tissue tumors: value of static and dynamic gadopentetate dimeglumine-enhanced MR imaging in prediction of malignancy. *Radiology*. 2004;233(2):493-502.
102. Fletcher BD, Hanna SL, Fairclough DL, Gronemeyer SA. Pediatric musculoskeletal tumors: use of dynamic, contrast-enhanced MR imaging to monitor response to chemotherapy. *Radiology*. 1992;184(1):243-248.
103. Shapeero LG, Vanel D, Verstraete KL, Bloem JL. Fast magnetic resonance imaging with contrast for soft tissue sarcoma viability. *Clin Orthop Relat Res*. 2002(397):212-227.
104. Verstraete KL, Dierick A, De Deene Y, et al. First-pass images of musculoskeletal lesions: a new and useful diagnostic application of dynamic contrast-enhanced MRI. *Magn Reson Imaging*. 1994;12(5):687-702.
105. Liu PT, Chivers FS, Roberts CC, Schultz CJ, Beauchamp CP. Imaging of osteoid osteoma with dynamic gadolinium-enhanced MR imaging. *Radiology*. 2003;227(3):691-700.
106. American College of Radiology. ACR Appropriateness Criteria®, Follow-up of Malignant or Aggressive Musculoskeletal Tumors. 2014; <http://www.acr.org/~media/ACR/Documents/AppCriteria/Diagnostic/FollowupMalignantOrAggressiveMusculoskeletalTumors.pdf>. Accessed October 8, 2014.
107. Hayashida Y, Yakushiji T, Awai K, et al. Monitoring therapeutic responses of primary bone tumors by diffusion-weighted image: Initial results. *Eur Radiol*. 2006;16(12):2637-2643.
108. Uhl M, Saueressig U, Koehler G, et al. Evaluation of tumour necrosis during chemotherapy with diffusion-weighted MR imaging: preliminary results in osteosarcomas. *Pediatr Radiol*. 2006;36(12):1306-1311.
109. Baur A, Stabler A, Bruning R, et al. Diffusion-weighted MR imaging of bone marrow: differentiation of benign versus pathologic compression fractures. *Radiology*. 1998;207(2):349-356.
110. Hayashida Y, Hirai T, Yakushiji T, et al. Evaluation of diffusion-weighted imaging for the differential diagnosis of poorly contrast-enhanced and T2-prolonged bone masses: Initial experience. *J Magn Reson Imaging*. 2006;23(3):377-382.
111. Maeda M, Matsumine A, Kato H, et al. Soft-tissue tumors evaluated by line-scan diffusion-weighted imaging: influence of myxoid matrix on the apparent diffusion coefficient. *J Magn Reson Imaging*. 2007;25(6):1199-1204.
112. van Rijswijk CS, Kunz P, Hogendoorn PC, Taminiau AH, Doornbos J, Bloem JL. Diffusion-weighted MRI in the characterization of soft-tissue tumors. *J Magn Reson Imaging*. 2002;15(3):302-307.
113. Ballinger JR, Kang H, Sweeney CA, Scott JD, Croker BP, Scott KN. P-31 changes as a measure of therapy response in resistant and sensitive osteosarcomas implanted into nude mice. *Magn Reson Imaging*. 1995;13(6):877-883.

114. Kettelhack C, Wickede M, Vogl T, Schneider U, Hohenberger P. ³¹P phosphorus-magnetic resonance spectroscopy to assess histologic tumor response noninvasively after isolated limb perfusion for soft tissue tumors. *Cancer*. 2002;94(5):1557-1564.
115. Millis K, Weybright P, Campbell N, et al. Classification of human liposarcoma and lipoma using ex vivo proton NMR spectroscopy. *Magn Reson Med*. 1999;41(2):257-267.
116. Moller HE, Vermathen P, Rummeny E, et al. In vivo ³¹P NMR spectroscopy of human musculoskeletal tumors as a measure of response to chemotherapy. *NMR Biomed*. 1996;9(8):347-358.
117. Singer S, Millis K, Souza K, Fletcher C. Correlation of lipid content and composition with liposarcoma histology and grade. *Ann Surg Oncol*. 1997;4(7):557-563.
118. Zakian KL, Shukla-Dave A, Meyers P, et al. Identification of prognostic markers in bone sarcomas using proton-decoupled phosphorus magnetic resonance spectroscopy. *Cancer Res*. 2003;63(24):9042-9047.
119. Fayad LM, Barker PB, Bluemke DA. Molecular characterization of musculoskeletal tumors by proton MR spectroscopy. *Semin Musculoskelet Radiol*. 2007;11(3):240-245.
120. Fayad LM, Barker PB, Jacobs MA, et al. Characterization of musculoskeletal lesions on 3-T proton MR spectroscopy. *AJR Am J Roentgenol*. 2007;188(6):1513-1520.
121. Fayad LM, Bluemke DA, McCarthy EF, Weber KL, Barker PB, Jacobs MA. Musculoskeletal tumors: use of proton MR spectroscopic imaging for characterization. *J Magn Reson Imaging*. 2006;23(1):23-28.
122. Oya N, Aoki J, Shinozaki T, Watanabe H, Takagishi K, Endo K. Preliminary study of proton magnetic resonance spectroscopy in bone and soft tissue tumors: an unassigned signal at 2.0-2.1 ppm may be a possible indicator of malignant neuroectodermal tumor. *Radiat Med*. 2000;18(3):193-198.
123. Schick F, Duda SH, Lutz O, Claussen CD. Lipids in bone tumors assessed by magnetic resonance: chemical shift imaging and proton spectroscopy in vivo. *Anticancer Res*. 1996;16(3B):1569-1574.
124. Wang CK, Li CW, Hsieh TJ, Chien SH, Liu GC, Tsai KB. Characterization of bone and soft-tissue tumors with in vivo ¹H MR spectroscopy: initial results. *Radiology*. 2004;232(2):599-605.
125. Sah PL, Sharma R, Kandpal H, et al. In vivo proton spectroscopy of giant cell tumor of the bone. *AJR Am J Roentgenol*. 2008;190(2):W133-139.
126. Baur-Melnyk A, Buhmann S, Becker C, et al. Whole-body MRI versus whole-body MDCT for staging of multiple myeloma. *AJR Am J Roentgenol*. 2008;190(4):1097-1104.
127. Dinter DJ, Neff WK, Klaus J, et al. Comparison of whole-body MR imaging and conventional X-ray examination in patients with multiple myeloma and implications for therapy. *Ann Hematol*. 2009;88(5):457-464.
128. Goo HW, Yang DH, Ra YS, et al. Whole-body MRI of Langerhans cell histiocytosis: comparison with radiography and bone scintigraphy. *Pediatr Radiol*. 2006;36(10):1019-1031.
129. Krohmer S, Sorge I, Krausse A, et al. Whole-body MRI for primary evaluation of malignant disease in children. *Eur J Radiol*. 2010;74(1):256-261.
130. Shortt CP, Gleeson TG, Breen KA, et al. Whole-Body MRI versus PET in assessment of multiple myeloma disease activity. *AJR Am J Roentgenol*. 2009;192(4):980-986.
131. Ma LD, Frassica FJ, Scott WW, Jr., Fishman EK, Zerbouni EA. Differentiation of benign and malignant musculoskeletal tumors: potential pitfalls with MR imaging. *Radiographics*. 1995;15(2):349-366.
132. Peh WC, Chan JH. Artifacts in musculoskeletal magnetic resonance imaging: identification and correction. *Skeletal Radiol*. 2001;30(4):179-191.
133. American College of Radiology. ACR Practice Parameter for Communication of Diagnostic Imaging Findings. 2014; <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/CommunicationDiag.pdf>. Accessed October 8, 2014.
134. Shellock FG. *Magnetic Resonance Procedures: Health Effects and Safety*. Boca Raton, Fla.: CRC Press; 2001.
135. Sawyer-Glover AM, Shellock FG. Pre-MRI procedure screening: recommendations and safety considerations for biomedical implants and devices. *J Magn Reson Imaging*. 2000;12(1):92-106.
136. American College of Radiology. ACR-AAPM Technical Standard for Diagnostic Medical Physics Performance Monitoring of Magnetic Resonance Imaging (MRI) Equipment. 2014; <https://www.acr.org/-/media/ACR/Files/Practice-Parameters/MR-Equip.pdf>. Accessed October 8, 2014.

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